GWAS 7

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GENOMIC RECOMBINATION



Sini Kerminen; www.helsinki.fi/~mjxpirin/stamp/

- Offspring inherits two genomes in continuous segments of the parent's two genomes
- Segments from recent ancestors are longer than from more distant ancestors
- This process determines correlations in GWAS data
 - Concept of genetic relatedness (last week's topic)
 - Sample-by-sample correlation
 - GWAS results at nearby SNPs are correlated (today's topic)
 - SNP-by-SNP correlation

HAPLOTYPES = HAPLOID GENOTYPES

I. True haplotypes



2. Observed (diploid) genotypes

A/G C/T A/C

I. Individual has inherited a chromosome with alleles A-T-C
from one parent and a chromosome with alleles
G-C-A at the same SNPs from the other parent.

These are the two **haplotypes** of the individual at these 3 SNPs.

2. Genotype data does not carry haplotype information for heterozygous loci:
We do not know whether A at SNP1 is coming from the same parent as C or as T at SNP2.

3. Possible haplotypes



3. **Haplotype phasing =** estimating which are the two haplotypes behind the observed diploid genotypes

EXAMPLE: 2500BPS REGION FROM CHR I

Southern Han Chinese

Finns

Luhya in Kenya

	Haplotypes			Haplotypes				Haplotypes										
rs115037027	т	т	т	С	т	т	т	С	т		т	т	т	т	С	т	т	т
rs 2409788	т	С	С	т	С	С	т	т	т		С	т	т	т	т	С	т	т
rs1576517	С	С	С	т	С	С	С	т	т		с	С	т	т	т	С	С	т
rs151240271	G	G	G	G	G	G	G	G	G		G	G	G	G	G	G	т	G
rs 2752436	т	т	т	т	т	т	т	т	С		т	т	т	С	т	т	С	т
rs76864380	т	т	т	А	т	т	т	А	т		т	т	т	т	А	т	т	А
rs6586443	G	G	А	G	G	А	G	G	G		G	G	G	G	G	А	G	G
rs35213023	G	G	G	G	G	G	G	G	А		G	G	G	А	G	G	А	G
rs34910942	С	С	С	С	С	С	С	С	G		с	С	С	G	С	С	С	С
Counts:	109	54	40	4	106	47	34	6	4		65	50	43	15	14	4	4	3
Freqs:	0.519	0.2571	0.1905	0.019	0.5354	0.2374	0.1717	0.0303	0.0202	0.3	8283	0.2525	0.2172	0.0758	0.0707	0.0202	0.0202	0.0152

From: LDHap https://Idlink.nci.nih.gov/?tab=Idhap

HAPLOTYPE BLOCK STRUCTURE



Haplotype is the sequence of alleles on the same chromosome, or, more generally, sequence of alleles inherited from the same parent.

SNP variation in the population is organized as haplotype blocks, where recombination seems to happen mainly between the blocks but little within any block.

Consequences for GWAS:

I. A relatively small number of tag-SNPs chosen from the blocks can capture most of (common) variation

2. It may be difficult to know which variant within a block is the causal variant as they are highly correlated



A 1.U



Recombination rate as function of physical position on MHC region shows 5 hotspots (triangles). Different colors are rates in different continental populations.

Most recombination events occur in a small proportion genome sequence.



Bherer. et al. 2017 Nature Communications 8, 14994

PHASING LEVERAGES SHARING OF HAPLOTYPE BLOCKS

Unphased genotypes	Possible p	hasing A	Possible p	ohasing B	Possible p	ohasing C	Possible	phasing D
A/C	А	С	А	С	А	С	А	С
G/T	G	Т	G	Т	Т	G	Т	G
A/T	А	Т	Т	А	A	Т	Т	А
Population haplotype frequency	55%	0%	15%	5%	2%	3%	0%	20%
Population frequency of 0% unordered haplotype pair		%	2 × (15% × 5%) = 1.5%		2 × (2% × 3%) = 0.12%		0%	
Posterior probability of unordered haplotype pair	0'	%	1.5% / (1.5% +	0.12%) = 93%	0.12% / (1.5%	+ 0.12%) = 7%	()%

Browning & Browning. (2011) Nature Reviews Genetics volume 12:703–714

Nature Reviews | Genetics

Consider one individual with a heterozygous genotype at each of three SNPs in a region. There are four possible haplotype configurations that are consistent with the genotype data (possible phasing patterns A–D). Suppose that haplotype frequencies are available from other individuals in the population at these sites (provided below each phasing pattern). The population frequency of a haplotype pair is obtained using the Hardy–Weinberg principle (independence of the two haplotypes within an individual); the factor of two in the frequency of the haplotype pairs accounts for both possible assignments of maternal and paternal origin to the two haplotypes. The posterior probabilities of the phased data are obtained from the population frequencies of the possible haplotype pairs. In this example, the posterior probability of phasing B (93%) is much greater than that of phasing C (7%).

EROSION OF HAPLOTYPE BACKGROUND OF A VARIANT



The mutation is indicated by a red triangle. Chromosomal stretches derived from the common ancestor of all mutant chromosomes are shown in yellow, and new stretches introduced by recombination are shown in blue. Markers that are physically close (that is, in the yellow regions of present-day chromosomes) tend to remain associated with the ancestral mutation even as recombination limits the extent of the region of association over time.

Exactly shared segment around the variant is shrinking over time among the carriers of the variant.

LINKAGE DISEQUILIBRIUM



a | At the outset, there is a polymorphic locus with alleles A and a. b | When a mutation occurs at a nearby locus, changing an allele B to b, this occurs on a single chromosome bearing either allele A or *a* at the first locus (A in this example). So, early in the lifetime of the mutation, only three out of the four possible haplotypes will be observed in the population. The *b* allele will always be found on a chromosome with the A allele at the adjacent locus. **c** | The association between alleles at the two loci will gradually be disrupted by recombination. **d** | This will result in the creation of the fourth possible haplotype and an eventual decline in LD among the markers in the population as the recombinant chromosome (*a*, *b*) increases in frequency.

	LDPAIR						
CEU (Central Europe) rs7837688 cbr8:128539360	rs4242382 chr8:12851757 A G 0 180	3 180 (0.909)	LVVK (Kenya) rs7837688	rs4242 chr8:128 A 40	2382 517573 G 139	179 (0.904)	D' is a normalized version of D that has maximum of I.
Haplotypes	16 2 16 182 (0.081) (0.919	18 (0.091) 198	chr8:128539360 T	5 45 (0.227)	14 153 (0.773)	19 (0.096) 198	
G_G: 180 (0.90 T_A: 16 (0.087 T_G: 2 (0.01) G_A: 0 (0.0)	09) D 1) R ² Chi-sc p-value	2: 1.0 2: 0.8791 1: 174.0659 2: <0.0001	<u>Haplotypes</u> G_G: 139 (0.7 G_A: 40 (0.20 T_G: 14 (0.07 T_A: 5 (0.025	702) 02) 71) 5)	<u>Stat</u> D': R ² : Chi-sq: p-value:	istics 0.0464 0.0008 0.1541 0.6946	

rs7837688(G) allele is correlated with rs4242382(G) allele rs7837688(T) allele is correlated with rs4242382(A) allele

rs7837688 and rs4242382 are in linkage equilibrium

From LDpair https://ldlink.nci.nih.gov/



DECAY OF LD IN HUMANS

Linkage disequilibrium was calculated around 10,000 randomly selected polymorphic sites in each population, having first thinned each population down to the same sample size (61 individuals). The plotted line represents a 5 kb moving average.

Finns have longest span of LD together with Asian populations followed by Europeans.

Africans have clearly smaller span of LD.

1000 Genomes. Nature 2015.

LD FRIENDS



LDproxy gives all other variants that are highly correlated with the target SNP.

Note how r^2 decays after recombination hotspots on either side of the variant.

LDproxy reported 3 variants in perfect LD $(r^2=1)$ with this SNP.

We call SNPs in high LD with each other as "LD-friends". Definition of "high" can vary.

https://ldlink.nci.nih.gov/



LDMATRIX



rs115037027 rs12409788 rs1576517 rs151240271 rs12752436 rs76864380 rs6586443 rs35213023 rs34910942 rs75047573 rs111549857 rs4847155 rs72683794 rs35748418 rs67818703 rs72683796 rs78376885 rs12760470 rs11185370 rs72683798 rs141938920 rs3811504 rs3811505 rs3811506 rs72685703 rs6664203 rs6698497 rs12021633

LD IN 4 GENES IN 2 FINNISH COHORTS

Genes: APOE CETP GCKR PCSK9

Cohorts:

NFBC= Northern Finland Birth Cohort 1966 (n~ 4000)

FINRISK (n~3000)

84NS

250

300

350

Note how similar LD is across these two Finnish cohorts.





b) CETP, C $_{\chi\chi}$







NFBC

SNPs

SNPg

150







Cichonska et al. 2016 Bioinformatics

MC4R - BMI REGION (LOCKE ET AL. 2015 NATURE)



SNPs having similar P-values are in high LD with each other. Their genotype data look almost the same and therefore the regression model gives them essentially the same results.

It is not clear based on these marginal P-values alone which (if any of these) variants are the causal ones.

Three lead SNPs have been chosen and LD w.r.t them is shown for their LD-friends

MARGINAL EFFECT AT A NON-CAUSAL SNP

Consider SNPs A and B of which B is causal and its allele I has effect size λ What is the marginal effect β at SNP A because of LD with SNP B ?

Haplotype	0 – 0	I – 0	0 – I	1 - 1
Frequency	f ₀₀	f ₁₀	f ₀₁	f _{II}
Effect	0	0	λ	λ

$$\begin{split} \beta &= E(Y \mid A = 1) - E(Y \mid A = 0) \\ &= \frac{f_{10}}{f_{10} + f_{11}} \times 0 + \frac{f_{11}}{f_{10} + f_{11}} \times \lambda - \frac{f_{00}}{f_{00} + f_{01}} \times 0 - \frac{f_{01}}{f_{00} + f_{01}} \times \lambda \\ &= \left(\frac{f_{11}}{f_{10} + f_{11}} - \frac{f_{01}}{f_{00} + f_{01}}\right) \times \lambda = \left(\frac{f_{11}}{f_A} - \frac{f_{01}}{1 - f_A}\right) \times \lambda \\ &= \left(\frac{f_{11}(1 - f_A) - f_{01}f_A}{f_A(1 - f_A)}\right) \times \lambda = \frac{f_{11} - (f_{11} + f_{01})f_A}{f_A(1 - f_A)} \times \lambda \\ &= \frac{f_{11} - f_B f_A}{\sqrt{f_A(1 - f_A)f_B(1 - f_B)}} \sqrt{\frac{f_B(1 - f_B)}{f_A(1 - f_A)}} \times \lambda = r_{AB} \sqrt{\frac{f_B(1 - f_B)}{f_A(1 - f_A)}} \lambda \end{split}$$

 f_A = allele I frequency at SNP A f_B = allele I frequency at SNP B r_{AB} = correlation of allele I at A and B

Conclusion:

The marginal effect at SNPA is shrunk towards 0 by correlation r_{AB} compared to SNP's B causal effect



Marginal effect at SNP A is a linear combination of the causal effects of all variants in LD with A, where the weights are the correlations with A (after scaling the genotypes).

$$\beta_A^* = \lambda_A^* + r_{A1} \lambda_1^* + r_{A2} \lambda_2^* + r_{A3} \lambda_3^* + r_{A4} \lambda_4^* + \dots$$

* denotes scaled effect: the allelic efect multiplied by $\sqrt{2f(1-f)}$, where f is MAF of the SNP

MASKING EFFECT BY LD

		rs768	7945		
		G	Α		
5220	G	1 20.8%	1.16 (1.04-1.29) 41.9%	1.11	1.26
rs35(Α	1.31 (1.17-1.47) 27.9%	1.64 (1.41-1.90) 9.4%	1.39	(1.16-1.37)
		1.18	1.25		
		1.0 (0.98-	07 -1.15)		

Viewing the data this way makes clear that the risk allele at the second SNP rs7687945 is more commonly found with the protective allele at rs356220 than would be expected were the SNPs in linkage equilibrium. As a result, the unconditional risk of the rs7687945 A allele (1.25) relative to the G allele (1.18) is 1.07 and not significantly different from 1.0

Shown in the centre of the table are estimates of odds ratio, 95% confidence limit (in brackets) and percentage frequency of the four haplotypes defined by the alleles at rs356220 and rs7687945. In the margins of the table is the risk of each of the alleles obtained by averaging the odds ratio of two haplotypes on which the allele can be found, weighting by the sample frequency. For example, the risk of carrying the G allele at rs356220 unconditional on the allele carried at rs7687945 is 1.11 (given in the top right) and is calculated as $(1 \times 20.8 + 1.16 \times 41.9)/(20.8 + 41.9)$.

By comparing the unconditional risks of the two alleles at each SNP, we recover the odds ratio estimated from a single SNP analysis.

UK Parkinson disease consortium & WTCCC2 Human Molecular Genetics,

Volume 20, Issue 2, 15 January 2011, Pages 345–353

STEPWISE FORWARD SEARCH

- Starts by conditioning on the lowest P-value
- Continues until no additional variant reaches pre-defined P-value threshold



Spain & Barrett 2016

STEPWISE FORWARD SEARCH

+ Informs about multiple causal variants accounting for LD

- Does not necessarily find the optimal configuration
- Completely ignores the uncertainty of the causal

configurations



Spain & Barrett 2016

FINE-MAPPING

- Assign probabilities for each variant that the variant is (one of) the causal variant(s)
- Done in a Bayesian framework with prior assumptions
 - Prior probability of each variant being a causal variant?
 - Default: Each variant is equally likely to be causal
 - Prior distribution of the non-zero effects of the causal variants?
 - Default: N(0, τ^2) (See GWAS 4 for how to set τ^2)

FINE-MAPPING ASSUMING I CAUSAL VARIANT

- If there is exactly one causal variant in the region and it is among the genotyped variants, then the posterior probability of being causal is proportional to the single-SNP marginal Bayes factor of association (ABF from GWAS4)
- This idea can be extended to fine-mapping each independent signal of the region after we have conditioned on the other signals in the region when we have computed the GWAS statistics (betas and SEs) that are used in calculating ABFs
- For multiple causal variants, we use methods such as FINEMAP or SuSiE

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> Bioinformatics, 32(10), 2016, 1493–1501 doi: 10.1093/bioinformatics/btw018 Advance Access Publication Date: 14 January 2016 Original Paper

Genetics and population analysis



Christian Benner

FINEMAP: efficient variable selection using summary data from genome-wide association studies

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Defended PhD thesis on FINEMAP in January 2019

• Define causal configuration γ as a binary vector for variants

$$\gamma = 0 0 1 0 0 1 0$$

• This configuration says that variants 3 and 7 are causal and the others have no effects.

• Define causal configuration γ as a binary vector for variants

$$\gamma = 0 0 1 0 0 1 0$$

- In total there are 2^p configurations on p variants, but we will assume that only sparse configurations are plausible, say those with < 10 causal variants per a region.
- Ultimate goal is to compute probability for each configuration, given the observed GWAS data in the region.

- Define causal configuration γ as a binary vector for variants
- Each causal variant picks its effect from $N(0, s^2)$

$$p(oldsymbol{\lambda}|oldsymbol{\gamma}) = \mathcal{N}\left(oldsymbol{0}, s^2oldsymbol{\Delta}_{oldsymbol{\gamma}}
ight)$$

Causal configuration
$$\gamma$$

$$1 0 0 0 0 0 0 0 0 0 0$$

$$\Delta_{\gamma} = \operatorname{diag}(\gamma) = \begin{bmatrix} 1 & & \\ 0 & & \\ & \ddots & \\ & & 0 \end{bmatrix}$$

- Define causal configuration γ as a binary vector for variants
- Each causal variant picks its effect from $N(0, s^2)$
- For each configuration compute the Bayes factor (BF), i.e., how well the configuration explains the data relative to the null model

$$BF_{\gamma} = \frac{P(DATA|\gamma)}{P(DATA|NULL)} = \frac{\mathcal{N}(\boldsymbol{z}|\boldsymbol{0}, \boldsymbol{R} + \boldsymbol{s}^{2}\boldsymbol{R}\,\boldsymbol{\Delta}_{\gamma}\,\boldsymbol{R})}{\mathcal{N}(\boldsymbol{z}|\boldsymbol{0},\boldsymbol{R})}$$

- Define causal configuration γ as a binary vector for variants
- Each causal variant picks its effect from $N(0, s^2)$
- For each configuration compute the Bayes factor (BF), i.e., how well the configuration explains the data relative to the null model
- By combining BFs with prior probabilities of configurations we get the posterior probabilities

$$p_{\gamma} = P(\gamma | \text{DATA}) \propto \text{prior}_{\gamma} \times BF_{\gamma}$$

FINEMAP ALGORITHM



- Collect configurations from a high probability region using Shotgun stochastic search (Hans et al. 2007)
 - Memorize BFs of all those configurations seen during the search
 - Stop once not much new probability mass is found
- Renormalize posteriors with respect to the configurations visited

Benner et al. 2016



FINEMAP

GWAS

15q21/LIPC association with HDL cholesterol





NATIONAL INSTITUTE FOR HEALTH AND WELFARE FINLAND

FINRISK STUDY 20000 indivduals



Christian Benner

Surakka et al. Nat. Genet. 2015

Inclusion of multiple ancestries drives improved fine-mapping of LDL-C levels in DMTN locus.



Association of the DMTN intron variant rs900776 with LDL-Cholesterol in the admixed African, European, or multi-ancestry meta-analysis (**a**) or with DMTN gene expression (**b**).

The region spanned by the 99% credible sets (assuming a single causal variant) are shown in the centre box. The LDL-C association signal significantly colocalizes with the expression signal of *DMTN* in liver.

c, The LD patterns for variants in the European ancestry 99% credible set differ greatly between African (AFR) and European ancestry individuals in 1000 Genomes. The lead variant has a posterior probability of 0.86 in the admixed African analysis, 0.51 in the European analysis and >0.99 in the multi-ancestry analysis.